

Critical Issues: The Caveolin Model

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Caveolin proteins

Homo sapiens



Caveolin-1 binds to cholesterol

Murata et al. (1995)

Sequence alignment: Caveolin - PITP α

The caveolin model

Possible hit in the database

Hit		Prob- ability	E-value	P-value
1	1kcm_A	PITP_alpha	20.2	0.00064
2	2jwa_A	ERB-2	19.5	0.0025

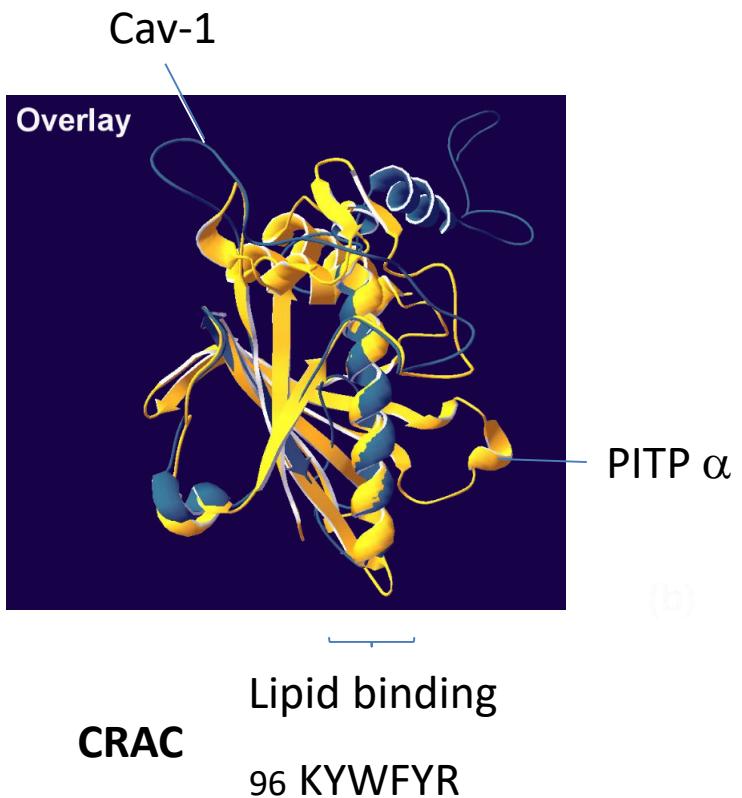
			Score	SS	Cols	Query	HMM
1	1kcm_A	PITP alpha	18.9	-0.3	160	1-178	
2	2jwa_A	ERB-2	15.2	3.5	44	73-149	
			Template	HMM			
			77-269	(270)			
			1-	44	(44)		

Fiedler (2008)

(see www.klausfiedler.ch/cav1pitp.pdf)

The only good scoring full-length alignment

Lipid exchange of Caveolin-1?



Previous models [Samhan-Arias et al. (2012)] could not be compared due to a lack of statistical parameters

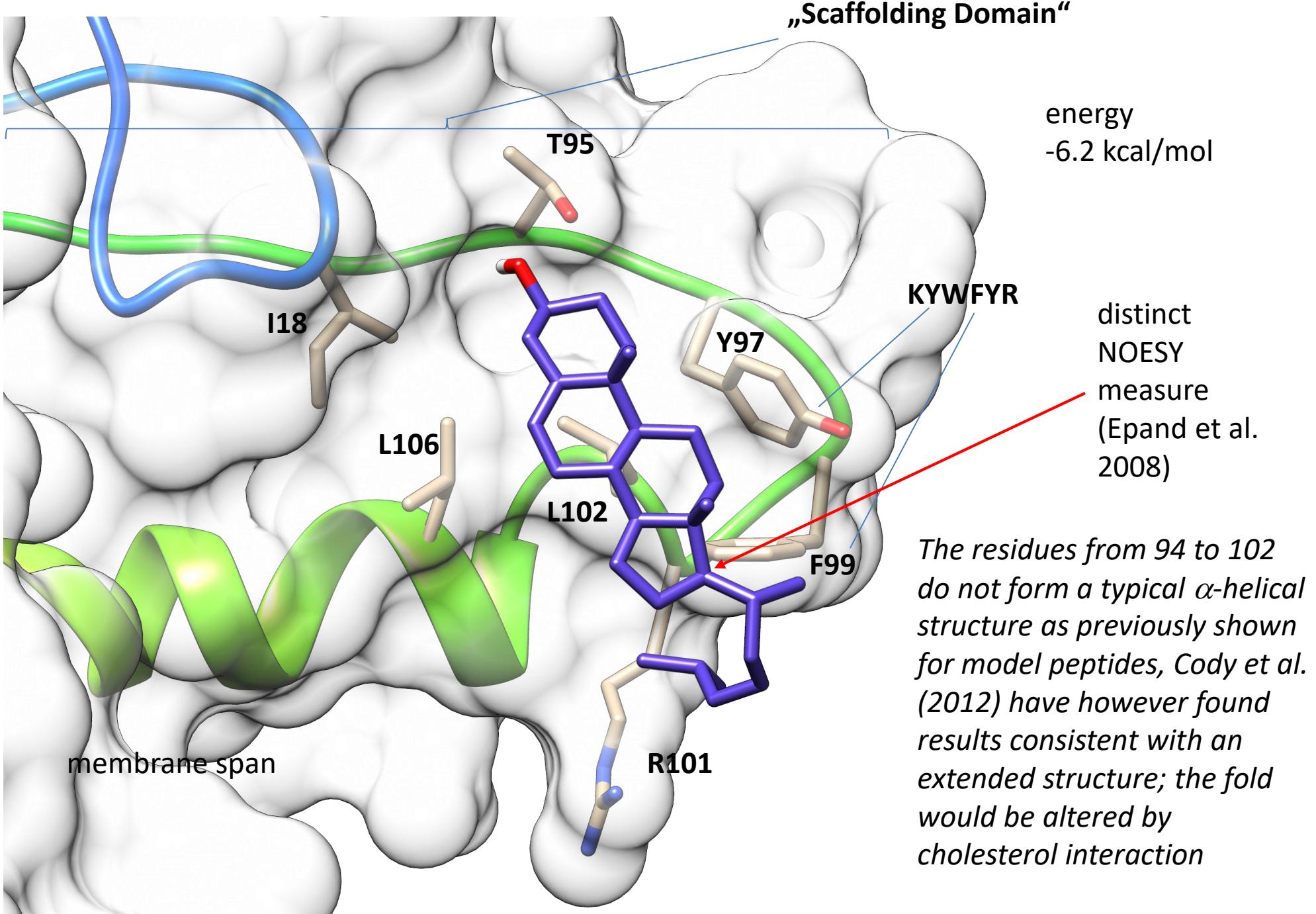
original model
based on
alignment

CRAC binding

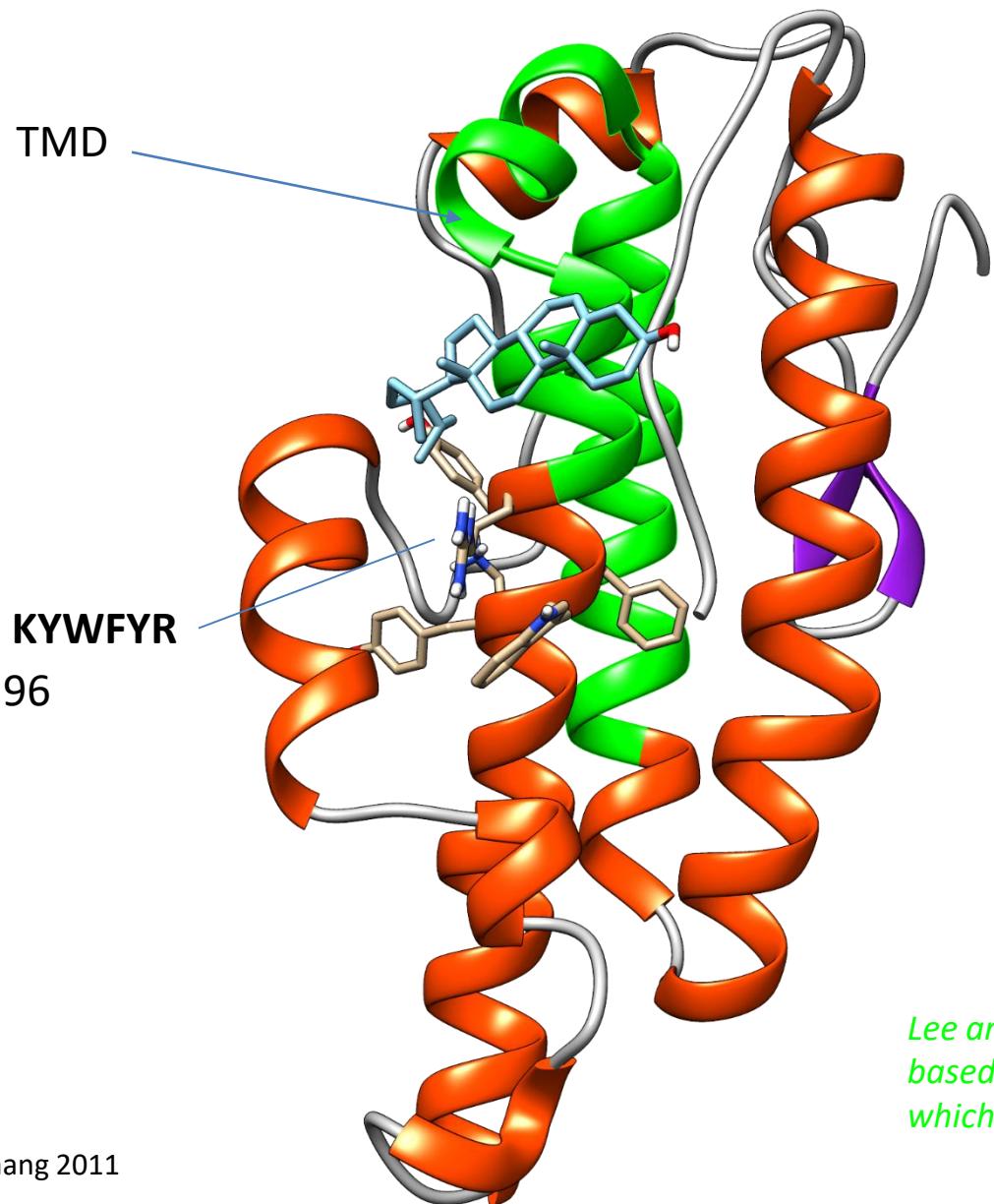
This model was generated with MODELLER and binding determined. Loop modeling was introduced to further optimize the structure (see Shen and Sali 2006) (www.klausfiedler.ch/Modeling_of_Caveolin.htm).

The distinct structure probability has not been determined since abundant hydrophobic residues preclude the easy use of folding energy functions. Statistical potentials were used (DOPE).

Distances
< 4 Å



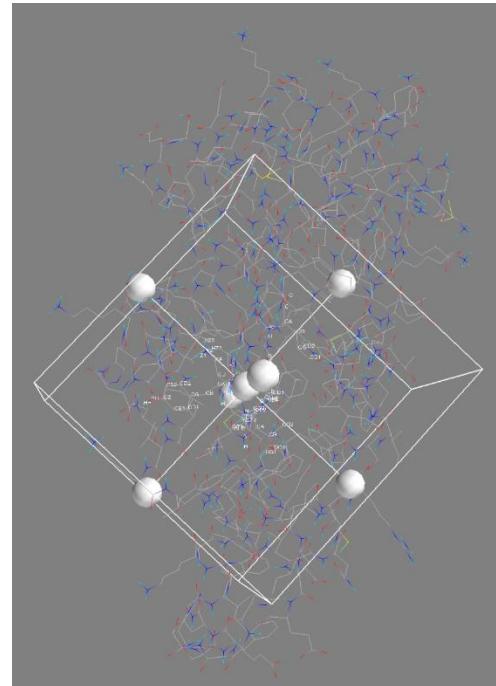
Ab initio predicted structures don't expose membrane residues



see
Xu and Zhang 2011

energy
 ≤ -5.9 kcal/mol

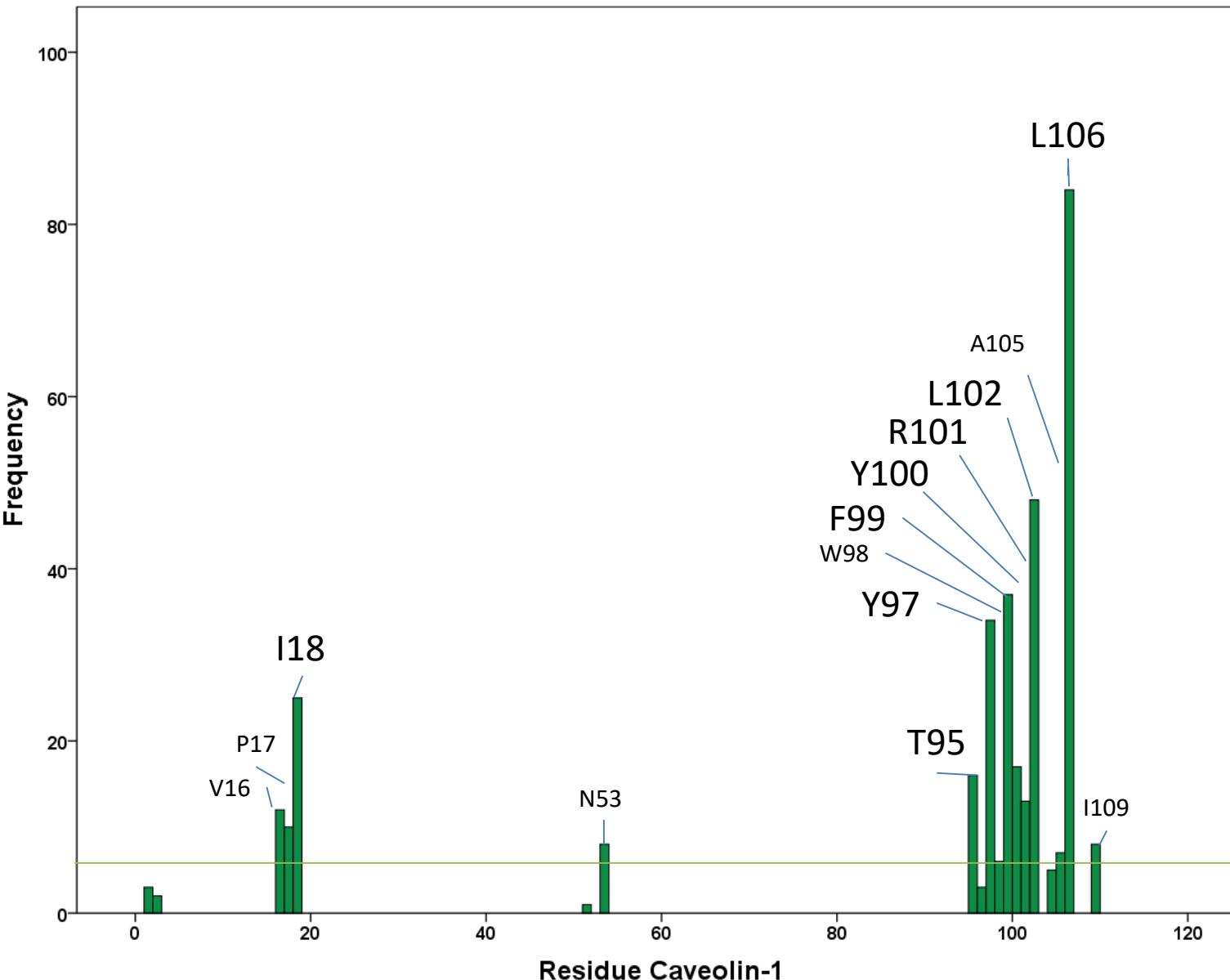
other stretches
in the cav1
structural
models may
have higher
affinity



PyRx docking to the CRAC residues,
cholesterol interacts distal to the CRAC
residues

*Lee and Glover 2012 predicted a helical membrane span
based on NMR analysis of a reconstituted cav1 96-136 peptide
which in this structure is prevalently shielded from solvent*

CRAC binding in the caveolin model: PITP α - homologue



Statistical evaluation
of CRAC binding of

cholesterol

cut-off Van der Waals
 $\geq -0.4 \text{ \AA}$
(9 models)

The cavatin effector stretch is located in region 89-95 of the CSD (cav1 scaffolding domain)(~82-101). The region distal to the effector stretch binds to cholesterol in the present model.

Mutants in caveolin proteins

Cav-1	Cav-3	Equivalent Cav-1	Conserved Cav1/2/3	Disease
Val14Leu		Val41		SIDS
Arg27Gln		Arg54	Yes	RMD/LGMD-1C/HyperCK
Asp28Glu		Asp55	Yes	RMD/LGMD-1C
Pro29Leu		Pro56	Yes	RMD/HyperCK/DM
Asn33Lys		Asn60	Yes	LGMD-1C/HyperCK
Lys38X		Lys65	Yes	LGMD-1C
Val44Glu		Val71	Yes	LGMD-1C/SIDS
Ala46Val/Thr		Ala73	Yes	LGMD-1C/RMD
Glu47Ala/Lys		Glu74	Yes	RMD
Ser53Gly		Ser80	Yes	RMD
Gly56Ser		Gly83		LGMD-1C
Val57Met		Ile84		HyperCKemia
Ser61Arg		Ser88	Yes	LGMD-1C/HyperCK
ΔThr64Phe65Thr66		91,92,93	*	LGMD-1C
Thr64Pro		Thr91		LGMD-1C
Trp71Ter		Trp98		RMD
Cys72Trp		Phe99		LGMD-1C
Thr78Met		Ala105		LQTS/SIDS/HyperCK/LGMD-1C
Phe107Leu	Leu79Arg	Phe107		SIDS/Breast Cancer
	Ala85Thr	Ala112	Yes	LQTS
	Leu87Pro [†]	Ile114		RMD
Gly116Ser		Gly116	Yes	Breast Cancer
	Ala93Thr [†]	Ala120	Yes	RMD/LGMD-1C
	ΔPhe97	Phe124		LGMD-1C/RMD/HyperCK
	Phe97Cys	Phe124		LQTS
Leu125Gln		Leu125		Breast Cancer
Pro132Leu	Pro105Leu	Pro132	Yes	LGMD-1C/Breast Cancer
Cys133Arg		Cys133		Breast Cancer
Ser136Cys		Ser136		Breast Cancer
Ile141Phe		Ile141		Breast Cancer
Tyr148His		Tyr148		Breast Cancer
	Ser141Arg	Ser168	Yes	LQTS

*Phe conserved

[†]labeled differently in Kubisch et al. (2003)
table adapted from Gazzero et al. (2010)

Cholesterol Interaction
CRAC
Putative Interaction

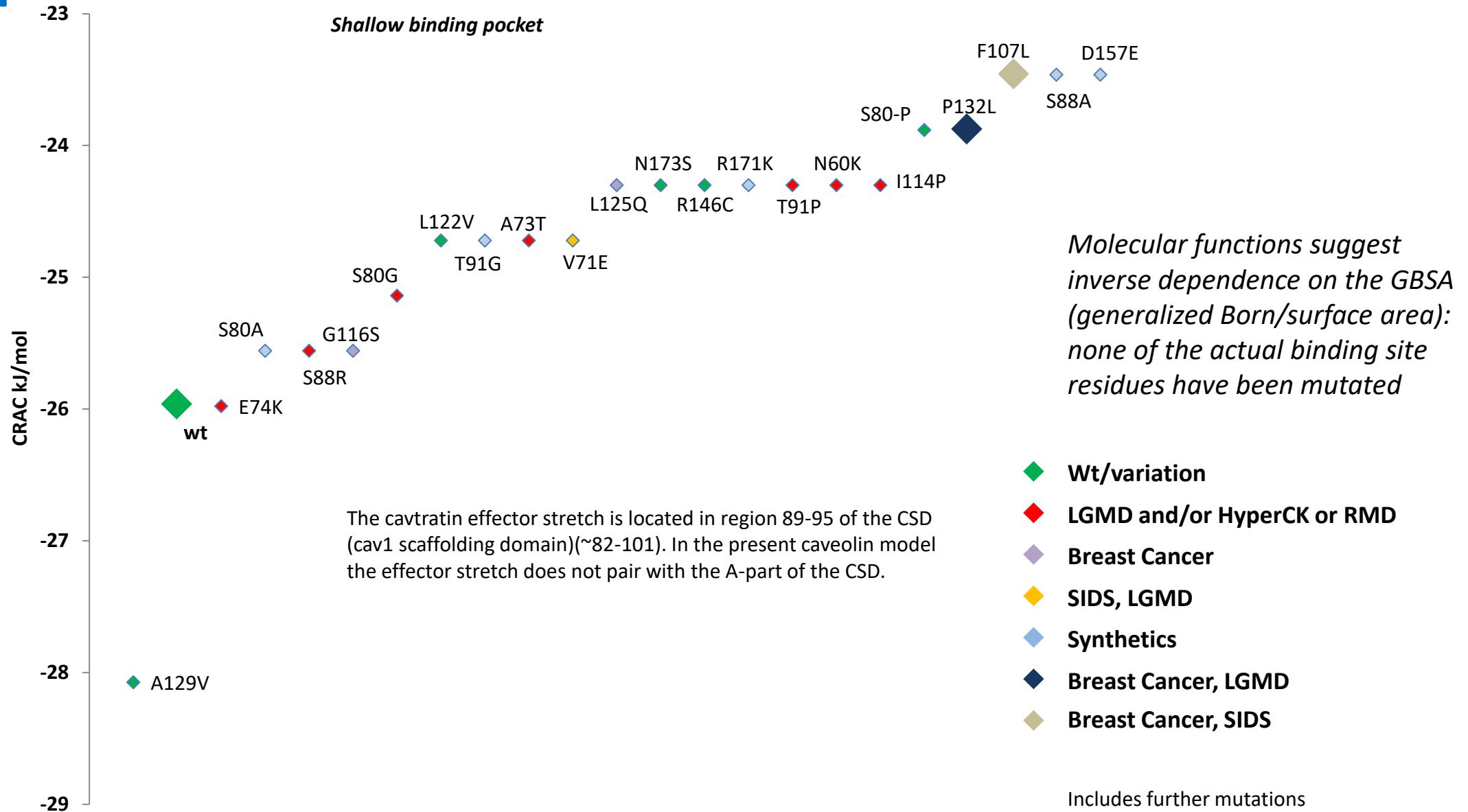
4 Å

cavratin-peptide
(see Bernatchez et al. 2005)



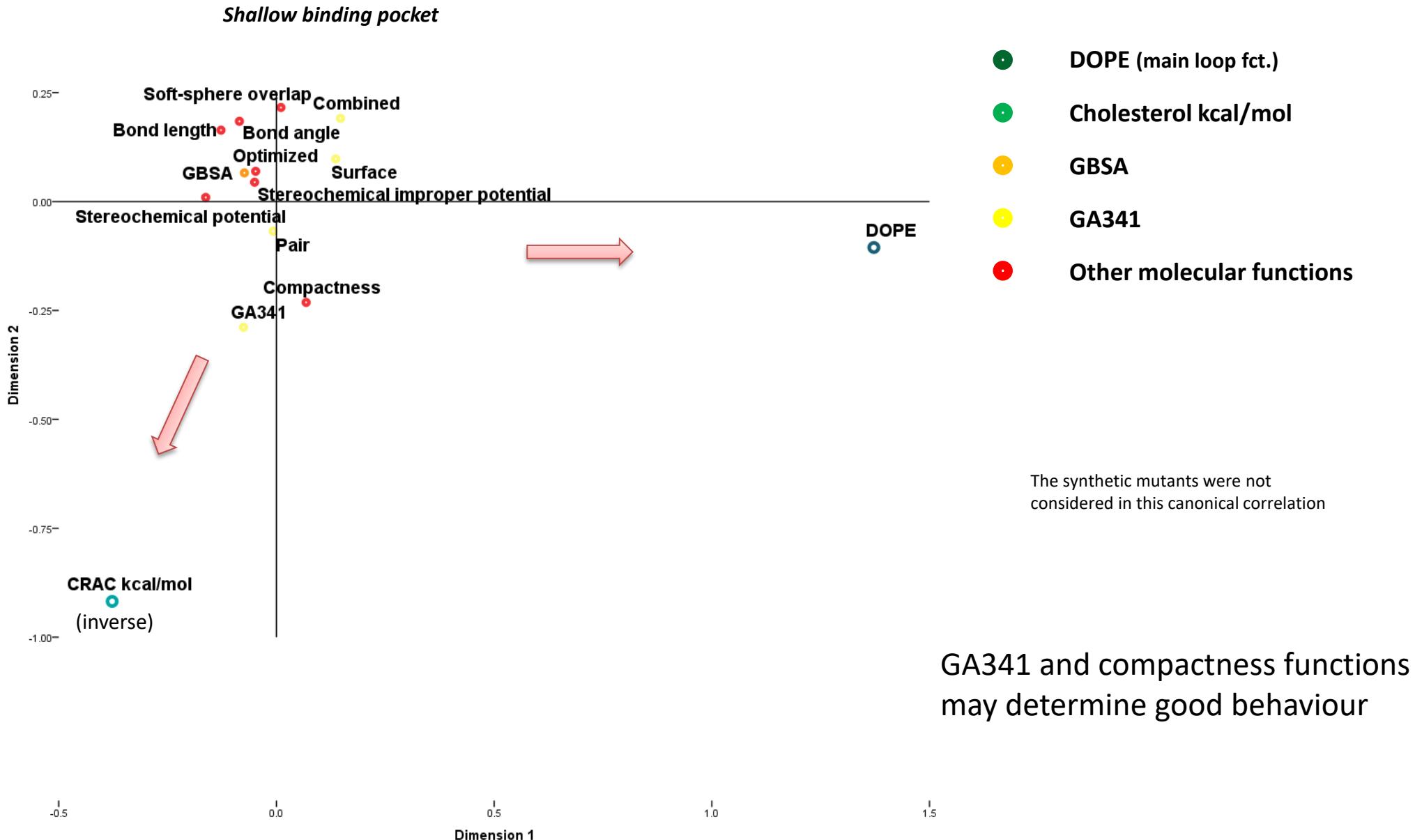
cavratin-peptide has
been used for eNOS
(endothelial nitric oxide
synthase) inhibition
studies

Mutations implicated in breast cancer and SIDS: Putative structural model

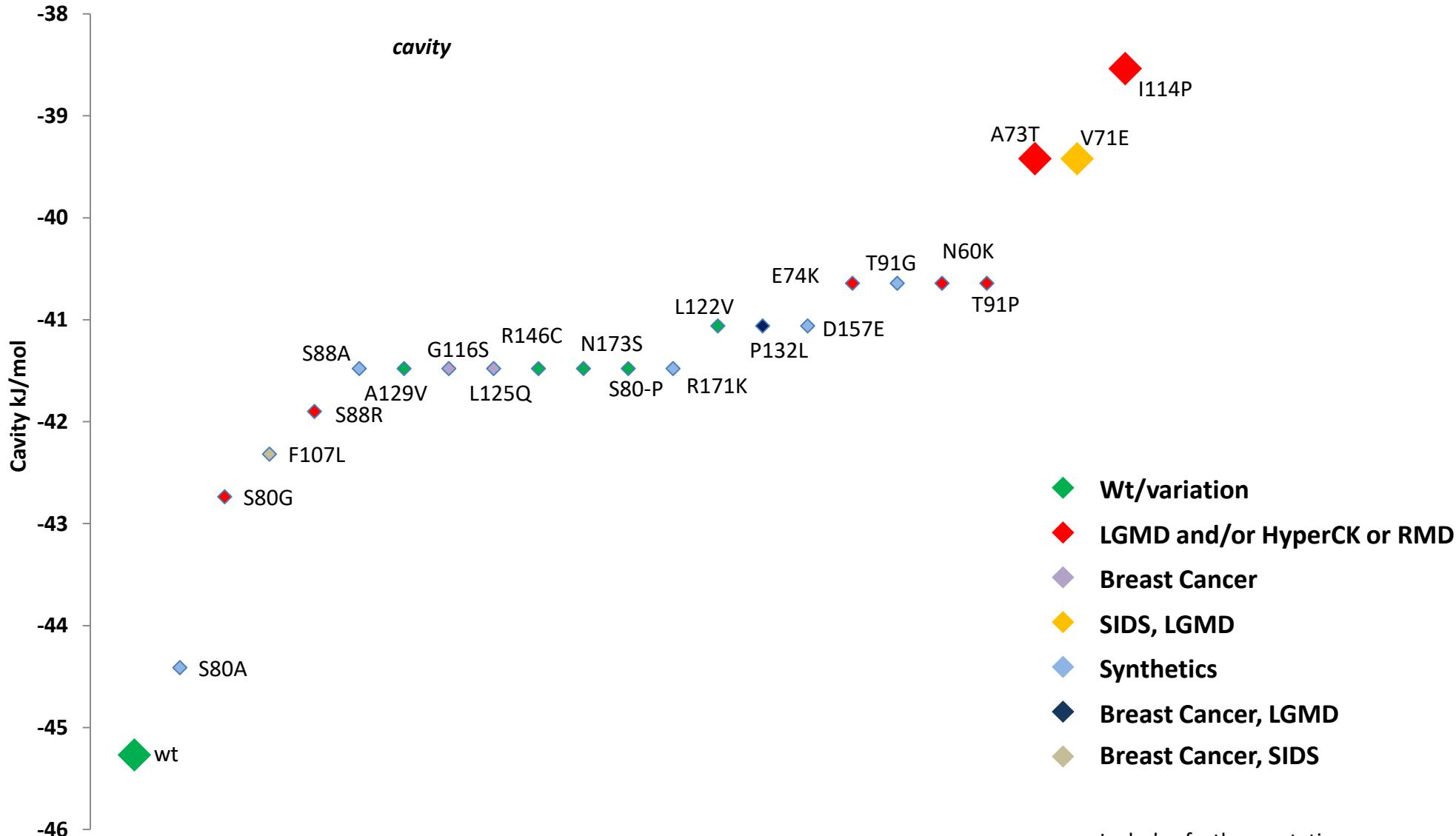


Folding in this series of cholesterol docking mutants

Overall canonical correlation

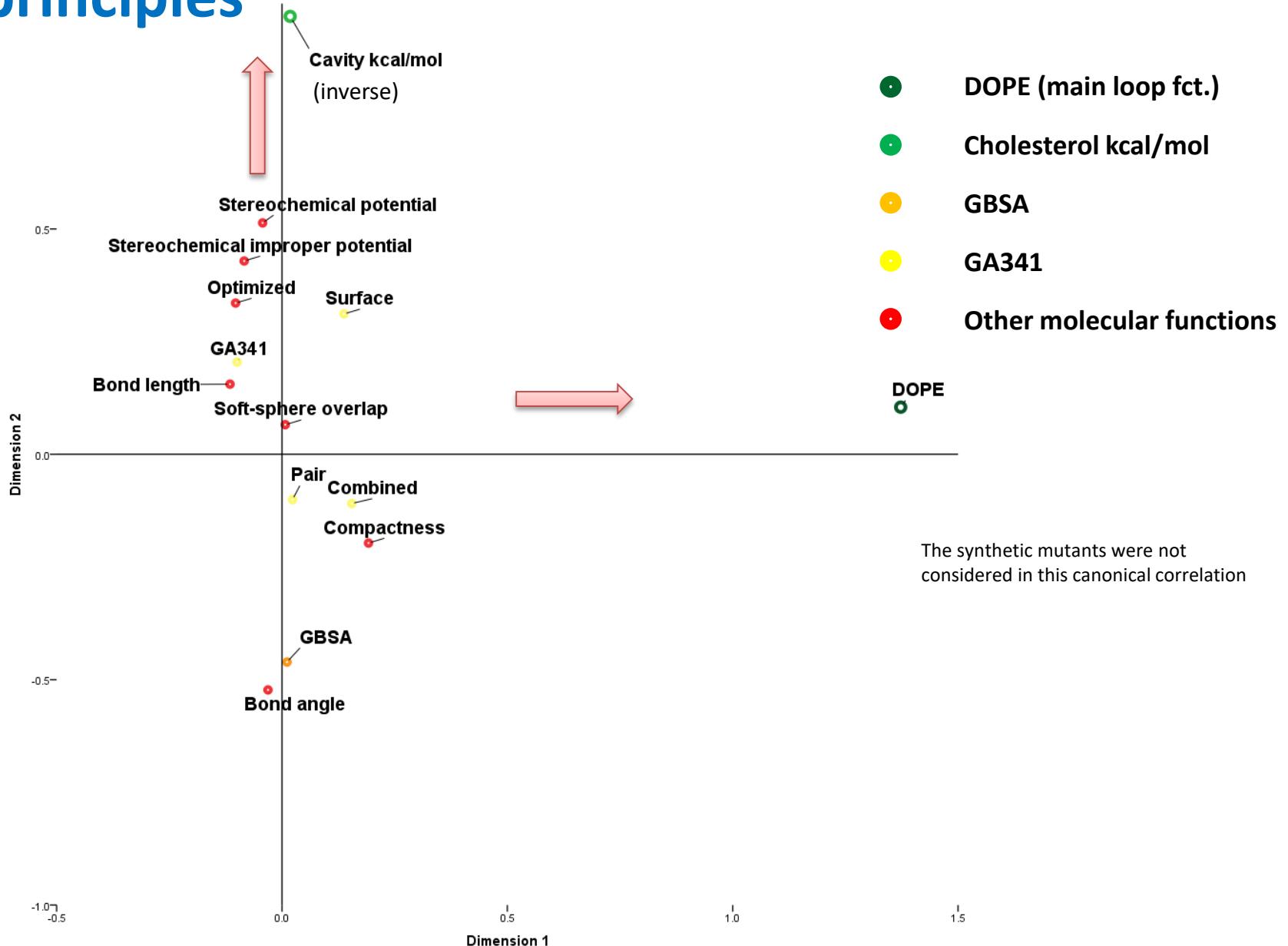


Mutations implicated in LGMD, RMD and SIDS: Putative cavity structural model



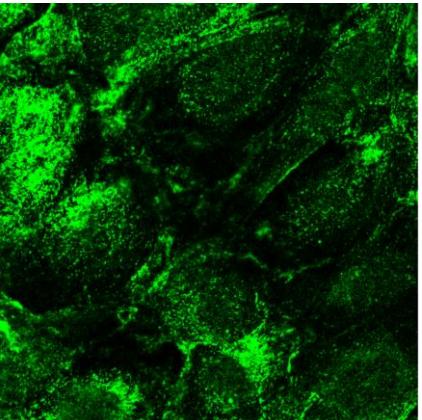
Hypothetical cavity interaction may be determined by different structural folding principles

Overall canonical correlation



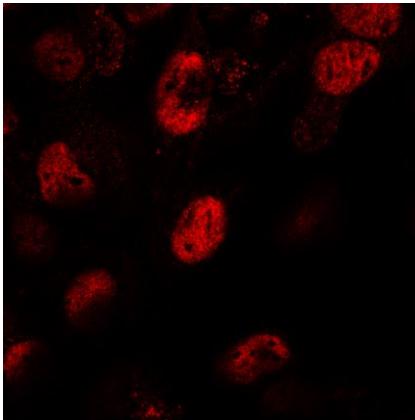
Nuclear caveolin-1 likely shows a particular conformer

Polyclonal antibodies (C13630)



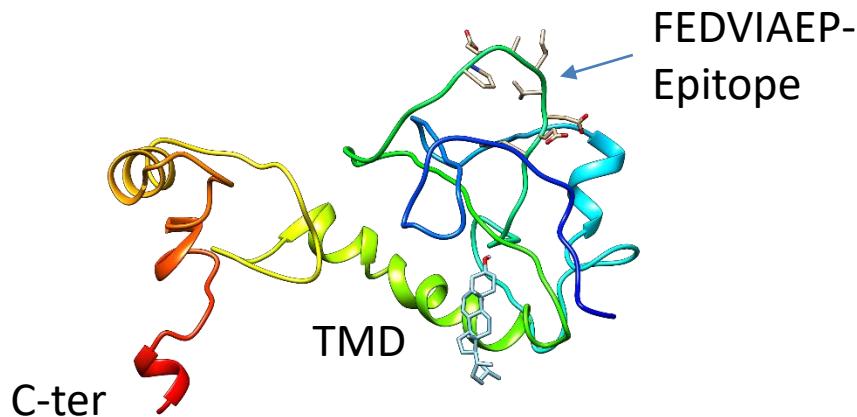
SVEC4-10

Polyclonal Signature-domain anti-bodies



SVEC4-10

The **folding** of caveolin may require the membrane bilayer and cholesterol and transport to the nucleus could, however, involve diffusion through the cellular cytoplasm wherein most hydrophobic residues are covered by amphipathic lipids and/or exposed to the cytoplasm as previously found for domains of intrinsically disordered proteins (Riback et al. 2017)



**Nuclear localization
with cav-1 antibodies**

Chatenay-Rivauday et al. 2004